

The Efficiency of Plant Growth Promoting Rhizobacteria for The Enhancement **of Rice Production**

1 2 3 4 4 2 Ravinder Polapally , Raj Bahadur , Raghu K , Kahkashan Parvin ,Aparna Srivastava , Neeraj Kumar

¹Department of Microbiology, Osmania University, Hyderabad, TS-India ²Department of Agronomy, Aacharya Narendra Deva University of Agriculture and Technology (ANDUA&T), Kumarganj, *Ayodhya-224229 (UP), India ³Department of Botany, Osmania University, Hyderabad, TS-India ⁴Department of Food and Nutrition, Era University, Lucknow-India*

Citation: Ravinder Polapally, Raj Bahadur, Raghu K, Kahkashan Parvin, Aparna Srivastava, Neeraj Kumar (2023). The Eficiency of Plant Growth Promoting Rhizobacteria for The Enhancement of Rice Production. Acta Botanica Plantae. V02i02, **53-57. DOI:** http://dx.doi.org/10.5281/zenodo.8340295

Corresponding Author: **Raj Bahadur |** E-Mail: **(drrajbahadur@nduat.org)**

Received 18 January 2023 | Revised 21 April 2023 | Accepted 11 June 2023 | Available Online June 24 2023

Copyright: This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Numerous bacteria encourage the growth of plants, and numerous microbial products that encourage plant growth have been *commercialized. In this review, we focus only on bacteria that originate from the root and have this impact on it. These microorganisms* are frequently referred to as PGPRs (plant-growth-promoting rhizobacteria). These rhizobacteria's beneficial impacts on Direct or *indirect plant growth are possible. The use of plant growth-promoting rhizobacteria (PGPR)* for agricultural purposes is becoming *more and more popular on a global scale, and it appears that this will be the future trend. PGPR are bioresources that have the potential* to revolutionize agriculture by offering significant advantages. These advantageous, free-living microorganisms promote emergence, *colonize roots, encourage growth, and increase yield. Many plant diseases are reported to be resistant to PGPR* in a variety of crops, including cereals, pulses, ornamentals, vegetables, plantation crops, spices, and some trees. The majority of studies have focused on the *investigation and possible advantages of PGPR in horticulture, forestry, and agriculture. Although well-documented, the possible mechanisms* used by these rhizobacteria in growth promotion and resistance have not yet been extensively investigated. When many *mechanisms are combined through the application of PGPR, their positive properties are increased. Due to inconsistent performance* and little commercialization outside of a few industrialized nations, their use hasn't been maximized. The benefits and drawbacks of using PGPR as bioinoculants, biofertilizers, and biocontrol agents, as well as their practical potential for bettering agriculture, are also *covered.* The goal of the study was to describe plant growth-promoting rhizobacteria (PGPR) for growth enhancement and stress tolerance features as well as their effectiveness on early establishment of rice seedlings. Out of 30 PGPR isolates, 18 fixed nitrogen, 17 solubilized tricalcium phosphate, and 29 and 17 generated IAA with or without the presence of L-tryptophane, according to in vitro growth boosting features When it came to stress tolerance, PGPR isolates were tolerant of pH levels between 5 and 10, NaCl levels between 1 and 6%, and polyethylene glycol (PEG) levels between 10 and 40%, respectively. With PIRG values ranging from 7 to 68%, *they* demonstrated antagonistic action against Pyriculariaoryzae.To maintain the development and yield of rice plants that are stressed by drought, it may be useful and environmentally benign to inoculate them with plant growth-promoting rhizobacteria *(PGPR).*

Keywords:Rice, Growth Promotion, Plants, Rhizobacteria

INTRODUCTION

More than half of the world's food needs are met by the cereal crop known as rice. More than 700× 106 tonnes of rice were produced worldwide in one year from 167 million tonnes of rice. Irrigated lowland rice provides more than 75% of the world's total rice production. In general, rice has been produced in looded ields with a constant water depth of 5 to 10 cm. Lowland rice is often directly seeded or transplanted on puddled soils by harrowing and leveling management after plowing under saturated water conditions [1-3]. A signiicant irrigation water supply is needed in looded areas, which is employed both as a management tool during rice cultivation and to meet the water requirements for the growth and development of rice plants [4]. It is well recognized that plants can beneit from plant growth-promoting rhizobacteria (PGPR) through a variety of direct and indirect processes, including biological nitrogen fixation, phosphate and potassium solubilization, synthesis of plant growth regulators, siderophore, hydrolyzing enzymes, and many more processes. By creating systemic resistance in plants, these helpful microbes also function as

biocontrol agents against pests and diseases [5]. Despite the overwhelmingly positive results, there are varying problems when using PGPR on rice plants. Since rice agriculture requires a large amount of N-fertilizer, the biological nitrogen fixation (BNF) process, which can reduce the use of chemical or inorganic N-fertilizer, is what has the greatest positive impact. According to estimates, rice crops remove 19.4 kg of nitrogen for every tonne of rice grain produced. Additionally, the regularly used N-fertilizer urea has a low plant uptake eficiency, frequently at only 40%. Despite the energy-intensive manufacturing procedures that use the non-renewable resource natural gas [6].

The legume-rhizobia symbiosis, in which the bacteria fix atmospheric nitrogen as endosymbionts inside root nodules in a nutrient-rich and oxygen-controlled microenvironment, has received the greatest attention from investigations of N2-fixing plant-microorganism interactions [7-8]. Rhizobia only modulates one host species during this symbiotic relationship, and there are hardly any cross-inoculations. Researchers have recently experimented with the idea of inoculating rhizobia on non-legumes like rice, and there have been sporadic reports of success, specifically through the improvement of rice seedling growth and grain output. The mechanisms, however, are yet unknown, and it was not the development of root nodules like in the rhizobia-legume symbiosis [9-10]. The researchers have hypothesized that this may be because rhizobia behaves and functions similarly to PGPR in delivering positive benefits such as phytohormone synthesis, phosphate solubilization, and an improvement in soil N assimilation [11]. Naturally contains a photosynthetic Bradyrhizobium sp. strain, which was isolated. Acetylene reduction assay (ARA) and greenhouse studies revealed that this strain produced a significant level of N2 fixing activity with 20% increases in shoot and grain yields. Contrarily, several other researchers who used ARA and 15N dilution methodologies in their experiments concluded that rhizobial inoculation had positive effects on rice that were more closely related to physiological adjustments in rice development and root shape than to BNF [12-13]. The use of PGPR and rhizobial strains as multiple-strain inocula for crops to profit from their various advantageous properties has recently attracted a lot of interest. A significant quantity of Nfertilizer can be replaced by BNF produced by several diazotrophic bacteria, such as Azotobacter, Clostridium, Azospirillum, Herbaspirillum, and Burkholderia, whereas Rhizobium can enhance the physiological growth or root morphology of rice plants [14-15]. This multi-strain biofertilizer inoculum is hypothesized to support plant growth and rice grain yield through BNF in addition to other known positive effects of PGPR and rhizobia [16].

This will increase the development and yield of the rice plants while reducing the need for artificial N-fertilizer. As a result, there will be a decrease in production costs, less environmental impact, and promotion of environmentally friendly and sustainable agriculture. The goal of the current study was to compare the effects of locally isolated PGPR and native rhizobia multi-strain biofertilizers on rice plant development and nitrogen fixation activities. Rice is regarded as a significant food and has an average yield of 4.5 t ha-1 every season, selfsuficiency is only about 75% [17-20]. Rice output needs to be raised by roughly 7 ha-1 per season to keep up with the rising demand. However, rice-based agricultural systems are

susceptible to climate change. Over 65% of the world's rice demand is met by vast stretches of rice cultivation in dangerous places, mostly along South and Southeast Asia's coastlines [21- 22]. Climate change causes the sea level to increase, resulting in flooding and the incursion of saltwater into inland areas. According to reports, more than 50% of arable land will be in danger by 2050 as a result of soil salinization, a side effect of climate change, incorrect irrigation methods, overuse of chemical fertilizers, and a lack of adequate drainage systems. Additionally, it is anticipated that by 2056, salt stress will damage around 100,000 ha of rice-growing regions [23].

Salinity negatively disrupts the physical and chemical qualities of soil and more severely influences crop growth. Beneficial microorganisms called plant growth-promoting rhizobacteria (PGPR) may be crucial in reducing the severity of this condition [24]. A beneficial substitute for inorganic fertilizers and pesticides, this species of rhizospheric bacteria can colonize plant roots and preserve soil fertility.PGPR is beneicial in boosting the development of a variety of crops when they are exposed to salt stress in prior studies. It has been reported that local strains are more effective at boosting plant resistance to salinity stress than PGPR derived from the non-saline ecosystem in salinity mitigation. The preliminary selection of locally-isolated salt-tolerant PGPR is crucial to ensure its effectiveness. Exopolysaccharide (EPS), an extracellular polymeric molecule that guarantees the survival of these beneficial microorganisms under unfavorable soil conditions, is one of the strategies these microbes use to mitigate salt stress [25-27].

Exopolysaccharides are also necessary for the growth of bacteria that aggregate or locculate, which is accomplished by the specific adsorption of the polymeric segment and the formation of polymer bridges between the cells. Additionally, EPS is useful for enhancing bacterial interaction and forming bacterial biofilms on plant root surfaces [28-29]. According to earlier research, several bacterial taxa, including Pseudomonas, Bacillus, Burkholderia, Enterobacter, Microbacterium, Planococcus, and Halomonas, are capable of producing EPS in conditions of salt stress. Additionally, PGPR can produce a variety of plant growth-promoting qualities, including the production of indole acetic acid, biological nitrogen fixation, the solubilization of soil phosphorus (P) and potassium (K), the production of siderophores, and the hydrolyzing of enzymes in salt-stressed conditions [30-32].

It has long been known that PGPR and plants go together. As endophytes that provide the plant host with growth-promoting traits, improve osmolyte, antioxidant, and phytohormonal signaling, and improve plant nutrient uptake eficiency, PGPR may also have some other potential mechanisms for coping with salinity. Not all PGPR can enter the plant host as endophytes, thus there are still more methods to investigate [33]. Rice output in Malaysia could be significantly increased by the use of salt-tolerant PGPR, particularly in the coastline region, which is subject to saltwater intrusion [34]. We postulate that choosing appropriate native and local bacterial strains and isolating them to reduce salt stress in rice plants through their several advantageous features would be a climate-smart agricultural practice and to characterize, identify, and ascertain the impact of salt-tolerant PGPR isolated from the saline rice field on crop growth and yield, the current study was carried out.

Technologies for Enhancing Rice Production

Production in Paddy Field: For the rice crop in paddy fields, several technologies have been created to reduce input costs and increase resource utilization eficiency. Some of these technologies are described since their use can increase yield while using less water.

Selection of suitable period of crop plantation:

- Rice farming recommendations included the following:
- 1. Plant during the rainy season and harvest after the season,
- 2. To determine the growing date for non-photoperiod sensitive types (110 to 120 days), the harvesting date should be determined after the rainy season,
- 3. Cropping schedules should be established according to the dry and wet seasons.

Increase yield per unit evapotranspiration

The new "IRRI varieties" have around a 3-fold increase in water productivity compared to the conventional kinds by boosting yield while concurrently reducing crop duration and the outflows of evapotranspiration, seepage, and percolation. New cultivars can be created and used to shorten agricultural seasons and use less input [35].

Laser leveling: Surface-irrigated fields have been leveled successfully using laser-guided machinery. It brings down the field's unevenness to around 2 cm, which improves water application and distribution eficiency, water productivity, fertilizer eficiency, and weed pressure. Up to 50% in savings on wheat, and 68% on rice. Therefore, the most useful innovation in agriculture is laser land leveling, notably for eficient and equitable input consumption for maximum output [36].

Direct seeding: When opposed to transplanting, direct seeding of rice saves time and energy. Dry soil seeding conserves water because there is no puddling and the entire time needed for plants to grow from seed to seed is cut down to roughly 10 days. The crops in the following rotation had higher yields and water eficiency. However, weed control in dry direct-seeded rice is more challenging than in puddled and transplanted rice. While typical transplanted rice on beds had a 15% reduction in total water input, direct-seeded rice had a reduction of 30% [37].

Better soil nutrient management: Although rice continues to require approximately the same quantity of water, better soil nutrient management leads to higher yields. Each kilogram of nitrogen fertilizer added to the soil could result in 10 to 15 kg more rice being produced [38].

Plant spacing: Each plant receives more room, air, and sunlight with broad spacing. Each plant thus produces more tillers. The roots would absorb more nutrients and develop robustly and widely. Since the plant is robust and vigorous, there will be more tillers [39]. The panicle contains more grains, and each grain weighs more as well 25×25 cm should be used as the rowto-row and plant-to-plant distances inside a row.

Weed control: Following the introduction of short-stature rice varieties and the practice of shallow water in the fields during the early stages, which created an ecological condition more conducive to their growth, weed management became even more dificult. A program for managing weeds that is both culturally and chemically sustainable is required. Chemicals

and cultural practices by themselves cannot effectively manage weeds. Without weed control, yield loss is predicted to be as high as 90% at a yield level of 7 to 8 tonnes per hectare [40]. The productivity of water is also increased with proper weed control.

Mulching and green manure: To preserve and improve soil fertility, mulching, and green manure are key methods of supplying organic matter to the soil. Crop wastes or green manure crops can be used as mulch, which offers food for soil life and mineral nutrients for plants. When legumes are utilized as green manure, they can add up to 200 kg/ha of nitrogen to the soil, which can save 50–75% of the mineral fertilizer needed for rice [41]. Mulch is applied on the soil's surface to prevent weed development, save water, protect against wind and water erosion, and reduce evaporation.

Climate and climatic change: Extreme weather occurrences like prolonged droughts, very high precipitation, and high temperatures have increased in frequency and intensity during the past few decades. Systems for agricultural production are quite vulnerable to these changes. Crop water needs can help with climate change adaptation by strengthening agricultural cropping systems' resilience and reducing their susceptibility to unfavorable climatic conditions [42]. Together or independently, these cutting-edge solutions have a positive influence on crop productivity and water savings. These methods increase soil fertility while simultaneously increasing productivity.

CONCLUSIONS

Reviewing earlier studies reveals that PGPRs have improved plant growth and yield in the greenhouse and in vitro environments by raising plant tolerance to non-biological stressors. However, these PGPRs frequently fail to produce these advantageous effects when utilized in natural settings, such as fields, which are frequently caused by insufficient rhizosphere and endorhizal colonization. The plant development and yield of several crops may be improved with PGPR. Despite these, it can raise agricultural output without harming the land or the environment by improving the quality of the crop's produce. Using effective soil and rhizome microorganisms in a soil-plant-environmental system allows for an agricultural system that is environmentally friendly, economically viable, and long-lasting.

In addition to providing safe rice and maintaining the health of the soil, the Purple Nonsulfur Bacteria (PNSB) mixture and P fertilizer decreased the amount of chemical fertilizer required for the maximum grain output. These methods are successfully used under the concept of advanced agriculture in many agricultural systems around the world without having a negative environmental impact and are universally adaptable to obtain the greatest benefits. Combining these many cuttingedge techniques, such as mulching, improved soil nutrient management, and direct sowing, leads to greater crop productivity and soil fertility in addition to water savings. The findings indicate that the rhizosphere is a significant source of microorganisms that can stimulate plant growth. Because PGPR bacteria can grow and function in environments with high levels of heavy metals, they may reduce the toxicity of heavy metals to plants. Due to the high ion toxicity, this impact is especially important in industrial regions without natural cover.

REFERENCES

- Giri, K., Mishra, G., Suyal, D. C., Kumar, N., Doley, B., Das, N., & Bora, N. (2023). Performance evaluation of native plant growth-promoting rhizobacteria for paddy yield enhancement in the jhum fields of Mokokchung, Nagaland, North East India. *Heliyon*, *9*(3). 1.
- Habib, S. H., Kausar, H., Saud, H. M., Ismail, M. R., & Othman, 2. R. (2016). Molecular characterization of stress tolerant plant growth promoting rhizobacteria (PGPR) for growth enhancement of rice. *Int. J. Agric. Biol, 18*, 184-191.
- Nandakumar, R., Babu, S., Viswanathan, R., Sheela, J., Raguchander, T., & Samiyappan, R. (2001). A new bioformulation containing plant growth promoting rhizobacterial mixture for the management of sheath blight and enhanced grain yield in rice. *Biocontrol*, *46*, 493- 510. 3.
- Cavite, H. J. M., Mactal, A. G., Evangelista, E. V., & Cruz, J. A. 4. (2021). Growth and yield response of upland rice to application of plant growth-promoting rhizobacteria. *Journal of Plant Growth Regulation*, *40*, 494-508.
- Sharma, A., Shankhdhar, D., & Shankhdhar, S. C. (2013). Enhancing grain iron content of rice by the application of plant growth promoting rhizobacteria. *Plant, Soil and Environment*, *59*(2), 89-94. 5.
- El-Mageed, A., Taia, A., El-Mageed, A., Shimaa, A., El-6. Saadony, M. T., Abdelaziz, S., & Abdou, N. M. (2022). Plant growth-promoting rhizobacteria improve growth, morphphysiological responses, water productivity, and yield of rice plants under full and deicit drip irrigation. *Rice*, *15*(1), 1-15.
- 7. Asiloglu, R., Shiroishi, K., Suzuki, K., Turgay, O. C., Murase, J., & Harada, N. (2020). Protist-enhanced survival of a plant growth promoting rhizobacteria, Azospirillum sp. B510, and the growth of rice (Oryza sativa L.) plants. *Applied Soil Ecology*, *154*, 103599.
- Mehmood, U., Inam-ul-Haq, M., Saeed, M., Altaf, A., Azam, F., 8. & Hayat, S. (2018). A brief review on plant growth promoting rhizobacteria (PGPR): a key role in plant growth promotion. *Plant protection*, $2(2)$, 77-82.
- Niranjan Raj, S., Shetty, H. S., & Reddy, M. S. (2006). Plant growth promoting rhizobacteria: potential green alternative for plant productivity. *PGPR: biocontrol and biofertilization*, 197-216. 9.
- Tan, K. Z., Radziah, O., Halimi, M. S., Khairuddin, A. R., & 10. Shamsuddin, Z. H. (2015). Assessment of plant growthpromoting rhizobacteria (PGPR) and rhizobia as multistrain biofertilizer on growth and N2 fixation of rice plant. *Australian Journal of Crop Science*, *9*(12), 1257-1264.
- 11. Vejan, P., Abdullah, R., Khadiran, T., Ismail, S., & Nasrulhaq Boyce, A. (2016). Role of plant growth promoting rhizobacteria in agricultural sustainability—a review. *Molecules*, *21*(5), 573.
- 12. Chen, D., Saeed, M., Ali, M. N. H. A., Raheel, M., Ashraf, W., Hassan, Z.,
- 13. Liu, Z., Zhang, X., Li, L., Xu, N., Hu, Y., Wang, C., & Li, D. (2022). Isolation and characterization of three plant growth-promoting rhizobacteria for growth enhancement of rice seedling. *Journal of Plant Growth Regulation*, *41*(3), 1382-1393.& Negm, S. (2023). Plant Growth Promoting Rhizobacteria (PGPR) and Arbuscular Mycorrhizal Fungi Combined Application Reveals Enhanced Soil Fertility and Rice Production. *Agronomy*, *13*(2), 550.
- 14. Datta, M., Palit, R., Sengupta, C., Pandit, M. K., & Banerjee, S. (2011). Plant growth promoting rhizobacteria enhance growth and yield of chilli ('Capsicum annuum'L.) under field conditions. *Australian Journal of Crop Science*, 5(5), 531-536.
- 15. Cavite, H. J. M., Mactal, A. G., Evangelista, E. V., & Cruz, J. A. (2021). Biochemical characteristics and inoculation effects of multi-trait plant growth-promoting rhizobacteria on upland rice (Oryza sativa L. cv PSB Rc23) seedling growth. *Archives of Microbiology*, 203(6), 3533-3540.
- Egamberdieva, D., Wirth, S., Bellingrath-Kimura, S. D., 16. Mishra, J., & Arora, N. K. (2019). Salt-tolerant plant growth promoting rhizobacteria for enhancing crop productivity of saline soils. *Frontiers in microbiology*, *10*, 2791.
- 17. de Andrade, L. A., Santos, C. H. B., Frezarin, E. T., Sales, L. R., & Rigobelo, E. C. (2023). Plant growth-promoting rhizobacteria for sustainable agricultural production. *Microorganisms*, *11*(4), 1088.
- 18. Bhattacharyya, P. N., & Jha, D. K. (2012). Plant growthpromoting rhizobacteria (PGPR): emergence in agriculture. *World Journal of Microbiology and Biotechnology*, *28*, 1327-1350.
- 19. Kong, Z., & Liu, H. (2022). Modification of rhizosphere microbial communities: A possible mechanism of plant growth promoting rhizobacteria enhancing plant growth and fitness. *Frontiers in Plant Science*, 13, 920813.
- 20. Vurukonda, S. S. K. P., Vardharajula, S., Shrivastava, M., & SkZ, A. (2016). Enhancement of drought stress tolerance in crops by plant growth promoting rhizobacteria. *Microbiological research*, *184*, 13-24.
- 21. Shultana, R., Kee Zuan, A. T., Yusop, M. R., & Saud, H. M. (2020). Characterization of salt-tolerant plant growthpromoting rhizobacteria and the effect on growth and yield of saline-affected rice. *PLoS One*, *15*(9), e0238537.
- 22. Enebe, M. C., & Babalola, O. O. (2018). The influence of plant growth-promoting rhizobacteria in plant tolerance to abiotic stress: a survival strategy. Applied microbiology and *biotechnology*, *102*, 7821-7835.
- 23. Arora, N. K., Fatima, T., Mishra, J., Mishra, I., Verma, S., Verma, R., & Bharti, C. (2020). Halo-tolerant plant growth promoting rhizobacteria for improving productivity and

remediation of saline soils. *Journal of Advanced Research*, *26*, 69-82.

- 24. Hayat, R., Ahmed, I., & Sheirdil, R. A. (2012). An overview of plant growth promoting rhizobacteria (PGPR) for sustainable agriculture. *Crop production for agricultural improvement*, 557-579.
- 25. Arriel-Elias, M. T., Oliveira, M. I., Silva-Lobo, V. L., Filippi, M. C. C., Babana, A. H., Conceição, E. C., & Cortes, M. D. C. (2018). Shelf life enhancement of plant growth promoting rhizobacteria using a simple formulation screening method.
- 26. Awlachew, Z. T., & Mengistie, G. Y. (2022). Growth Promotion of Rice (Oryza sativa L.) Seedlings Using Plant Growth-Promoting Rhizobacteria (PGPR) Isolated from Northwest Ethiopia. *Advances in Agriculture*, *2022*.
- 27. Rais, A., Shakeel, M., Hafeez, F. Y., & Hassan, M. N. (2016). Plant growth promoting rhizobacteria suppress blast disease caused by Pyricularia oryzae and increase grain yield of rice. *BioControl*, *61*, 769-780.
- 28. Ramamoorthy, V., Viswanathan, R., Raguchander, T., Prakasam, V., & Samiyappan, R. (2001). Induction of systemic resistance by plant growth promoting rhizobacteria in crop plants against pests and diseases. *Crop protection*, *20*(1), 1-11.
- 29. Paul, D., & Lade, H. (2014). Plant-growth-promoting rhizobacteria to improve crop growth in saline soils: a review. *Agronomy for sustainable development*, *34*, 737- 752.
- 30. Hoque, M. N., Hannan, A., Imran, S., Paul, N. C., Mondal, M. F., Sadhin, M. M. R., & Rhaman, M. S. (2023). Plant growthpromoting rhizobacteria-mediated adaptive responses of plants under salinity stress. *Journal of Plant Growth Regulation*, *42*(3), 1307-1326.
- 31. de Souza, R., Beneduzi, A., Ambrosini, A., Da Costa, P. B., Meyer, J., Vargas, L. K., & Passaglia, L. M. (2013). The effect of plant growth-promoting rhizobacteria on the growth of rice (Oryza sativa L.) cropped in southern Brazilian fields. *Plant and soil*, *366*, 585-603.
- 32. Hussain, M. B., Shah, S. H., Matloob, A., Mubaraka, R., Ahmed, N., Ahmad, I., & Jamshaid, M. U. (2022). Rice Interactions with Plant Growth Promoting Rhizobacteria. In *Modern Techniques of Rice Crop Production* (pp. 231- 255). Singapore: Springer Singapore.
- Saberi Riseh, R., Ebrahimi-Zarandi, M., Tamanadar, E., 33. Moradi Pour, M., & Thakur, V. K. (2021). Salinity stress: Toward sustainable plant strategies and using plant growth-promoting rhizobacteria encapsulation for reducing it. *Sustainability*, *13*(22), 12758.
- 34. Verma, M., Mishra, J., & Arora, N. K. (2019). Plant growthpromoting rhizobacteria: diversity and applications. *Environmental biotechnology: for sustainable future*, 129- 173.
- 35. Ojuederie, O. B., Olanrewaju, O. S., & Babalola, O. O. (2019). Plant growth promoting rhizobacterial mitigation of drought stress in crop plants: Implications for sustainable agriculture. *Agronomy*, *9*(11), 712.
- 36. Kumawat, K. C., Nagpal, S., & Sharma, P. (2022). Potential of plant growth-promoting rhizobacteria-plant interactions in mitigating salt stress for sustainable agriculture: A review. *Pedosphere*, *32*(2), 223-245.
- 37. Xiao, A. W., Li, Z., Li, W. C., & Ye, Z. H. (2020). The effect of plant growth-promoting rhizobacteria (PGPR) on arsenic accumulation and the growth of rice plants (Oryza sativa L.). *Chemosphere*, *242*, 125136.
- 38. Vimal, S. R., Patel, V. K., & Singh, J. S. (2019). Plant growth promoting Curtobacterium albidum strain SRV4: an agriculturally important microbe to alleviate salinity stress in paddy plants. *Ecological Indicators*, *105*, 553-562.
- 39. Dey, R., Pal, K. K., & Tilak, K. V. B. R. (2014). Plant growth promoting rhizobacteria in crop protection and challenges. In *Future challenges in crop protection against fungal pathogens*(pp. 31-58). New York, NY: Springer New York.
- 40. Dey, R. K. K. P., Pal, K. K., Bhatt, D. M., & Chauhan, S. M. (2004). Growth promotion and yield enhancement of peanut (Arachis hypogaea L.) by application of plant growth-promoting rhizobacteria. *Microbiological research*, *159*(4), 371-394.
- 41. Paungfoo-Lonhienne, C., Redding, M., Pratt, C., & Wang, W. (2019). Plant growth promoting rhizobacteria increase the eficiency of fertilisers while reducing nitrogen loss. *Journal of environmental management*, *233*, 337-341.
- 42. Hafeez, F. Y., Abaid-Ullah, M., & Hassan, M. N. (2013). Plant growth-promoting rhizobacteria as zinc mobilizers: a promising approach for cereals biofortification. In *Bacteria in agrobiology: crop productivity* (pp. 217-235). Berlin, Heidelberg: Springer Berlin Heidelberg.